

European Journal of Pharmacology 440 (2002) 173-187



Neuropeptide Y receptors as targets for anti-obesity drug development: perspective and current status

Eric Parker^{a,*}, Margaret Van Heek^a, Andrew Stamford^b

^aDepartment of CNS and Cardiovascular Research, Schering-Plough Research Institute, Mail Stop K-15-2-2760, 2015 Galloping Hill Road, Kenilworth, NJ 07033, USA ^bDepartment of Chemical Research, Schering-Plough Research Institute, 2015 Galloping Hill Road, Kenilworth, NJ 07033, USA

Received 3 December 2001; accepted 10 December 2001

Abstract

Neuropeptide Y is a widely distributed neuropeptide that elicits a plethora of physiological effects via interaction with six different receptors (Y_1-y_6) . Recent attention has focused on the role of neuropeptide Y in the regulation of energy homeostasis. Neuropeptide Y stimulates food intake, inhibits energy expenditure, increases body weight and increases anabolic hormone levels by activating the neuropeptide Y Y_1 and Y_5 receptors in the hypothalamus. Based on these findings, several neuropeptide Y Y_1 and Y_5 receptor antagonists have been developed recently as potential anti-obesity agents. In addition, mice lacking neuropeptide Y, the neuropeptide Y Y_1 receptor or the neuropeptide Y Y_5 receptor have been generated. The data obtained to date with these newly developed tools suggests that neuropeptide Y receptor antagonists, particularly neuropeptide Y Y_1 receptor antagonists, may be useful anti-obesity agents. However, the redundancy of the neurochemical systems regulating energy homeostasis may limit the effect of ablating a single pathway. In addition, patients in whom the starvation response is activated, such as formerly obese patients who have lost weight or patients with complete or partial leptin deficiency, may be the best candidates for treatment with a neuropeptide Y receptor antagonist. © 2002 Elsevier Science B.V. All rights reserved.

Keywords: Neuropeptide Y; Obesity; Neuropeptide receptor, neuropeptide Y; Anti-obesity agent

1. Introduction

Neuropeptide Y is a 36-amino acid neuropeptide that was discovered in 1982 in a directed search for C-terminally amidated peptides (Tatemoto et al., 1982). Neuropeptide Y is a member of the pancreatic polypeptide family; other members of this family are the structurally related peptides, peptide YY and pancreatic polypeptide (Tatemoto et al., 1982). Neuropeptide Y is widely distributed in both the central and peripheral nervous systems and is one of the most abundant neuropeptides known. In the periphery, neuropeptide Y is localized in post-ganglionic sympathetic neurons, adrenal medulla, enteric neurons, cardiac non-sympathetic neurons, certain non-adrenergic perivascular neurons and parasympathetic neurons (Grundemar and Hakanson, 1993; McDermott et al., 1993). In sympathetic neurons and adrenal medulla, the peptide is co-localized with the classical sympathetic neurotransmitter noradrenaline (Lundberg et al.,

1982). In the brain, neuropeptide Y-containing neuronal cell bodies are found primarily in the locus coeruleus, the nucleus of the solitary tract and the arcuate nucleus of the hypothalamus. These neuropeptide Y-containing neuronal cell bodies also typically contain other neurotransmitters such as noradrenaline and send projections throughout the brain; hence, neuropeptide Y can be found in most brain regions, particularly in the cortex, hippocampus, thalamus, hypothalamus and brainstem (Chronwall et al., 1985).

Since it was one of the first neuropeptides to be discovered and is ubiquitously distributed, a great deal is now known about the biology of neuropeptide Y. Neuropeptide Y has been implicated in a wide variety of physiological effects; hence, neuropeptide Y and its receptors have sparked a great deal of basic research and drug development interest. This review will focus on the role played by neuropeptide Y and neuropeptide Y receptors in the regulation of energy homeostasis. The reader is referred to other excellent reviews for a more comprehensive discussion of the biology of neuropeptide Y (Cerda-Reverter and Larhammar, 2000; Grundemar and Bloom, 1997; Wettstein et al., 1995).

^{*} Corresponding author. Tel.: +1-908-740-7389; fax: +1-908-740-3294. E-mail address: eric.parker@spcorp.com (E. Parker).

2. Neuropeptide Y receptors

Neuropeptide Y, peptide YY and pancreatic polypeptide elicit their physiological effects by interacting with at least six distinct G protein-coupled receptors designated Y₁, Y₂, Y₃, Y₄, Y₅ and y₆ (Michel et al., 1998). With the exception of the neuropeptide Y Y₃ receptor, genes and/or cDNAs encoding each of these neuropeptide Y receptors have been cloned. In contrast to other families of G protein-coupled receptors, the neuropeptide Y receptors share only modest primary sequence homology (30-50%). In fact, some neuropeptide Y receptors are structurally more related to G protein-coupled receptors outside of the neuropeptide Y receptor family. The structural differences among neuropeptide Y receptors is beneficial to drug discovery efforts since compounds with high affinity for a particular neuropeptide Y receptor are less likely to interact with other neuropeptide Y receptors. In addition to having distinct amino acid sequences, each of the neuropeptide Y receptors is characterized by a unique pharmacological profile (Table 1) and a distinct tissue localization.

2.1. The neuropeptide $Y Y_1$ receptor

The neuropeptide Y Y₁ receptor was initially identified in early pharmacological studies as the neuropeptide Y receptor responsible for contraction of isolated vascular preparations (Grundemar and Bloom, 1997). In these studies, the neuropeptide Y Y₁ receptor was pharmacologically defined by its high affinity for neuropeptide Y and [Pro³⁴]-peptide YY and its low affinity for N-terminally truncated neuropeptide Y analogues (Table 1). Neuropeptide Y Y₁ receptor mRNA is abundantly expressed in many rat and human brain regions, including hypothalamic centers controlling energy homeostasis (Caberlotto et al., 1997; Parker and Herzog, 1999). Neuropeptide Y Y₁ receptor binding

Table 1
Affinities of neuropeptide Y (NPY), peptide YY (PYY), pancreatic polypeptide (PP) and related peptides for the known neuropeptide Y receptors

Peptide	EC ₅₀ (nM)					
	rY ₁	rY ₂	rY ₃	rY ₄	rY ₅	my ₆
hNPY	0.14	1.2	1.8	>1000	0.96	1.9
PYY	0.70	0.58	7200	>1000	1.0	0.8
pNPY-(2-36)	3.4	1.6		>1000	1.2	1.4
pNPY-(3-36)	110	2.4		>1000	2.8	3.9
pNPY-(13-36)	300	2.2		>1000	20	16.1
[Pro ³⁴] PYY	0.37	>1000		6.0	1.3	
hPP	150	>1000		0.037	1.4	>1000
rPP	>1000	>1000		0.060	170	>1000

Data are from Gerald et al. (1996), with the exception of the rat neuropeptide Y Y_3 receptor (rY_3) data which are from Higuchi et al. (1988) and the mouse neuropeptide Y y_6 receptor (my_6) data, which are from Mullins et al. (2000). Prefixes before peptide or receptor names are as follows: h=human, p=porcine, r=rat, m=mouse.

sites are also abundantly expressed in many rat brain regions, including the hypothalamus (Caberlotto et al., 1998b; Dumont et al., 1996). In contrast, levels of neuropeptide Y Y₁ receptor binding sites are very low throughout most of the human brain, perhaps due to instability of the receptor during the post-mortem interval (Caberlotto et al., 1997). In the periphery, neuropeptide Y Y₁ receptor mRNA is expressed primarily in rodent kidney, heart, spleen, skeletal muscle, lung, gastro-intestinal tract and vascular smooth muscle (Goumain et al., 1998; Nakamura et al., 1995), and in human kidney, heart, lung, colon, testis, adrenal gland, placenta, bone marrow and vascular smooth muscle (Wharton et al., 1993).

2.2. The neuropeptide $Y Y_2$ receptor

The neuropeptide Y Y₂ receptor was also identified in early pharmacological studies as the neuropeptide Y receptor responsible for regulation of noradrenaline release from sympathetic nerve terminals (Grundemar and Bloom, 1997). The neuropeptide Y Y₂ receptor is pharmacologically unique in that it has high affinity for N-terminally truncated neuropeptide Y analogues and low affinity for [Pro34]peptide YY (Table 1). The neuropeptide Y Y2 receptor is localized in several rat and human brain regions, including hypothalamic nuclei regulating energy homeostasis (Caberlotto et al., 1998a; Parker and Herzog, 1999). The neuropeptide Y Y2 receptor is localized on neuropeptide Ycontaining neurons in the brain, suggesting that this receptor is an autoreceptor (Caberlotto et al., 2000; King et al., 2000). In the periphery, the neuropeptide Y Y₂ receptor can be pharmacologically detected on the terminals of rat sympathetic, parasympathetic and sensory neurons, again functioning as an autoreceptor or heteroreceptor (Grundemar and Bloom, 1997). Negligible levels of neuropeptide Y Y₂ receptor mRNA are generally found in peripheral tissues (Gehlert et al., 1996), although low expression of neuropeptide Y Y2 receptor mRNA has been reported in human and rat gastro-intestinal tract (Gehlert et al., 1996; Goumain et al., 1998).

2.3. The neuropeptide $Y Y_3$ receptor

The neuropeptide Y Y₃ receptor was originally characterized pharmacologically in bovine adrenal chromaffin cells and has subsequently been found in rat heart, brainstem, hippocampus, colon and lung (for references, see Michel et al., 1998). The distinguishing pharmacological feature of the Y₃ receptor is its much higher affinity for neuropeptide Y than for peptide YY (Table 1). Although pharmacological evidence supports the existence of the neuropeptide Y Y₃ receptor in rats and cows, there are no reports of this receptor in humans and the receptor has not yet been cloned. Cloning of this receptor will be required to unequivocally validate its existence and determine its physiological role.

2.4. The neuropeptide $Y Y_4$ receptor

The neuropeptide Y Y₄ receptor was initially identified by molecular cloning (Lundell et al., 1995). This receptor is pharmacologically characterized by its high affinity for pancreatic polypeptide and its low affinity for neuropeptide Y (Table 1; Lundell et al., 1995). Therefore, the neuropeptide Y Y₄ receptor is probably a pancreatic polypeptide receptor rather than a neuropeptide Y receptor. Neuropeptide Y Y₄ receptor mRNA is sparsely expressed in brain; expression is seen primarily in the brainstem, but also at low levels in other brain regions such as the hypothalamus (Parker and Herzog, 1999). In the periphery, the neuropeptide Y Y₄ receptor is expressed primarily in rat testis and lung (Lundell et al., 1996). A different pattern of peripheral expression is seen in humans, with expression primarily detected in colon, small intestine, prostate and pancreas (Lundell et al., 1996).

2.5. The neuropeptide $Y Y_5$ receptor

The neuropeptide Y Y₅ receptor was also initially identified by molecular cloning (Gerald et al., 1996). The neuropeptide Y Y₅ receptor is pharmacologically distinguished by its high affinity for both N-terminally truncated analogues of neuropeptide Y and [Pro³⁴]peptide YY (Table 1). In addition, the neuropeptide Y Y₅ receptor has high affinity for human pancreatic polypeptide, but much lower affinity for rat pancreatic polypeptide (Table 1). Neuropeptide Y Y₅ receptor mRNA is discretely localized in rat and human brain, primarily in piriform cortex, olfactory tubercle and hypothalamus (Nichol et al., 1999; Parker and Herzog, 1999). Neuropeptide Y Y₅ receptor binding sites have also been detected in these regions, although some groups fail to detect neuropeptide Y Y₅ receptor binding in hypothalamus (Dumont et al., 1998; Statnick et al., 1998; Widdowson et al., 1997a). Interestingly, neuropeptide Y Y₅ receptor mRNA is almost always localized in neurons that also express neuropeptide Y Y₁ receptor mRNA (Naveilhan et al., 1998). In the periphery, neuropeptide Y Y₅ receptor mRNA has been detected in rodent testis, spleen, pancreas, gastro-intestinal tract, vascular smooth muscle cells and cardiomyocytes (Gerald et al., 1996; Pellieux et al., 2000; Statnick et al., 1998). Neuropeptide Y has also been reported to alter diuresis, natriuresis and plasma glucose levels via neuropeptide Y Y₅ receptor activation in rats (Bischoff and Michel, 1999).

2.6. The neuropeptide $Y y_6$ receptor

The neuropeptide Y y_6 receptor was also initially identified by molecular cloning (Weinberg et al., 1996). A functional neuropeptide Y y_6 receptor gene is found in mice and rabbits, but a neuropeptide Y y_6 receptor gene has not been detected in rats (Burkhoff et al., 1998). The neuropeptide Y y_6 receptor gene is a non-functional pseudogene in rabbits and primates (Burkhoff et al., 1998). The neuropeptide Y y_6 receptor has a pharmacological profile that is similar to that

of the neuropeptide Y Y_1 receptor, but is somewhat more tolerant of N-terminal truncation (Table 1). In mice, neuropeptide Y y_6 receptor mRNA has been detected in kidney, testis and brain, particularly in the hypothalamus (Burkhoff et al., 1998; Weinberg et al., 1996). Although the neuropeptide Y y_6 receptor clearly does not contribute to the physiological effects of neuropeptide Y in humans, this receptor must be taken into account when considering physiological effects of neuropeptide Y in mice.

2.7. Peptide YY-preferring receptor

A putative neuropeptide Y receptor known as the peptide YY-preferring receptor has been characterized in several tissues. This receptor has approximately 5- to 10-fold higher affinity for peptide YY than for neuropeptide Y and was initially found in crypt cells in the epithelium of the rat small intestine (Laburthe et al., 1986). Recently, Goumain et al. (2001) provided convincing evidence that the peptide YY-preferring receptor in rat small intestine is in fact the neuropeptide Y Y_2 receptor. Thus, the peptide YY-preferring receptor can now be equated with the neuropeptide Y Y_2 receptor.

3. Physiological role of neuropeptide Y in energy homeostasis

3.1. Effect of exogenous neuropeptide Yon energy intake and energy expenditure

Neuropeptide Y has potent effects on a variety of behavioral, physiological and endocrine systems that are critical in the modulation of energy homeostasis. Neuropeptide Y is the most potent or exigenic peptide identified to date. Nearly two decades ago, it was discovered that administration of exogenous neuropeptide Y directly to the brains of rats causes a tremendous increase in food consumption, even under conditions of satiation (Clark et al., 1984; Stanley and Leibowitz, 1984). Injection of neuropeptide Y directly into specific brain nuclei shows that the peptide is most effective in stimulating feeding when administered in the perifornical hypothalamus (Stanley et al., 1993). Hence, neuropeptide Ycontaining neuronal pathways in the hypothalamus are most critical in the regulation of energy homeostasis. Central administration of neuropeptide Y to rats also decreases energy expenditure by decreasing sympathetic nervous system activity; as a result, thermogenic activity in brown adipose tissue, a key regulator of energy expenditure in rodents, is diminished (Egawa et al., 1991). By increasing food consumption and decreasing energy expenditure, central administration of neuropeptide Y results in a state of positive energy balance that will promote weight gain if chronically maintained. Central administration of neuropeptide Y also induces hyperinsulinemia, hyperglucagonemia, increased plasma non-esterified fatty acids and insulin resistance. These effects are independent of the hyperphagic effect of neuropeptide Y (Marks and Waite, 1996).

Chronic central administration of neuropeptide Y to normal rats results in many of the physiological abnormalities observed in the obese state, including hyperphagia, accelerated weight gain, increased adiposity, hypertriglyceridemia, hyperinsulinemia and hypercorticosteronemia (Vettor et al., 1994; Zarjevski et al., 1993). As observed with other obesity models, neuropeptide Y-induced obesity is glucocorticoiddependent, as adrenal ectomy prior to chronic central neuropeptide Y administration prevents this obesity syndrome from developing (Sainsbury et al., 1997), and dexamethasone treatment of adrenalectomized rats restores the response to chronic neuropeptide Y administration (Zakrzewska et al., 1999). Hence, chronic central administration of neuropeptide Y results in obesity that is strikingly reminiscent of spontaneous, genetic and diet-induced obesity in rodents and man. Transgenic mice modestly overexpressing neuropeptide Y have significantly increased body weight, plasma glucose and plasma insulin compared to wild type mice when maintained on a palatable high sucrose diet, although not when maintained on a low fat diet (Kaga et al., 2001). This effect could be partially explained by significant but transient hyperphagia immediately following introduction of the high sucrose diet.

3.2. Role of endogenous neuropeptide Y in the modulation of energy homeostasis

A large body of evidence also suggests that endogenous neuropeptide Y plays a central role in energy homeostasis. Immunoneutralization of hypothalamic neuropeptide Y decreases feeding, even in rats deprived of food for 24 h (Shibasaki et al., 1993). Administration of neuropeptide Y antisense oligodeoxynucleotides to the brains of rats leads to the expected decrease in neuropeptide Y levels in the arcuate nucleus and also significantly reduces natural feeding behavior (Akabayashi et al., 1994; but see also Flynn et al., 1999). Furthermore, administration of certain potent neuropeptide Y receptor antagonists to rodents decreases food intake and body weight (see below).

Consistent with a role in the tonic modulation of energy homeostasis, expression of neuropeptide Y mRNA and the synthesis of neuropeptide Y are altered with changes in nutritional state and metabolic need, as well as in a number of genetic and diet-induced models of obesity. Depriving lean rats of food for 48 h leads to a significant increase in neuropeptide Y mRNA expression in the arcuate nucleus, and an increase in neuropeptide Y itself in the arcuate nucleus and the paraventricular nucleus (Sahu et al., 1988). Thus, increased neuropeptide Y synthesis and release may mediate the hyperphagic response observed after fasting. Streptozotocin-induced diabetic rats, a model of insulindependent diabetes with high metabolic demand, have increased neuropeptide Y mRNA expression in the arcuate nucleus and increased neuropeptide Y levels and neuro-

peptide Y release in the paraventricular nucleus, which may in part be responsible for the hyperphagia exhibited by these animals (Sahu et al., 1990). The obese *ob/ob* mouse, which lacks the hormone leptin, and the obese db/db mouse and Zucker rat, which do not have functional leptin receptors, both exhibit increased levels of hypothalamic neuropeptide Y mRNA and peptide (Sanacora et al., 1990; Stephens et al., 1995). Thus, increased neuropeptide Y transmission may be partially responsible for the hyperphagia and obesity that are characteristic of these mice (see below). A number of investigators have also assessed changes in the neuropeptide Y system in diet-induced obese rodents, but no single conclusion can be made from these studies, perhaps due to the myriad of differences between experiments. Suffice it to note that there appears to be a dysregulation of the neuropeptide Y system in animals that are prone to develop obesity under the appropriate dietary conditions (e.g., Bergen et al., 1999; Guan et al., 1998; Levin and Dunn-Meynell, 1997; Wilding et al., 1992).

Considering the wealth of data indicating that the neuropeptide Y system is important in energy homeostasis, results from transgenic mice in which the neuropeptide Y system has been manipulated have been somewhat paradoxical. Mice that are deficient in neuropeptide Y have normal food intake and body weight, and display the same hyperphagia as their wild type counterparts after food deprivation (Erickson et al., 1996a). Further studies indicated that plasma corticosterone, insulin, and glucose levels were normal in neuropeptide Ydeficient mice (Erickson et al., 1997). A lack of neuropeptide Y also did not affect the development of obesity induced by diet or chemical means (Hollopeter et al., 1998). Studies by a separate group, however, demonstrated that neuropeptide Ydeficient mice have decreased food intake after a 24-48 h fast (Bannon et al., 2000). The conflicting data on the effect of neuropeptide Y deficiency on energy homeostasis in mice is puzzling in view of the wealth of data supporting a role for the peptide in energy homeostasis. However, the results may suggest that mice readily compensate during development for the absence of a key neurotransmitter. Indeed, neuropeptide Y deficient mice are more sensitive to the hyperphagic effect of agouti-related protein, a melanocortin receptor antagonist that is co-expressed with neuropeptide Y in arcuate neurons (see below; Marsh et al., 1999b). Furthermore, many neurotransmitter and neuropeptide knockout mice do not display overt phenotypes; phenotypes are frequently seen only under specific physiological conditions or when the organism is stressed in some way. In this regard, it is interesting to note that the obese phenotype of ob/ob mice is attenuated when neuropeptide Y is absent, suggesting that neuropeptide Y is partially responsible for this obesity syndrome (Erickson et al., 1996b). These data also implicate neuropeptide Y as a downstream mediator of leptin action in the brain (see below). In contrast to the *ob/ob* mouse, the obesity characteristic of the genetically obese UCP-DTA or Ay mice was not affected when these mice were crossed with neuropeptide Y deficient mice (Hollopeter et al., 1998).

3.3. Integration of neuropeptide Y with other brain pathways regulating energy homeostasis

In addition to neuropeptide Y, many other neurotransmitter and neuropeptide systems in the brain have been shown to influence energy homeostasis (for a review, see Spiegelman and Flier, 2001). Clearly, these manifold systems must interact with one another and the outputs of each of these systems must be highly integrated to achieve tight control of energy homeostasis. Consistent with its key role in the regulation of energy homeostasis, neuropeptide Y has been shown to interact with numerous neurotransmitter and neuropeptide systems that are thought to play a role in this process.

Leptin, a hormone secreted by adipose tissue that is critical in the regulation of body weight, exerts many of its physiological effects on body weight regulation by acting on target neurons in the brain (Spiegelman and Flier, 2001). It is likely that leptin is a key messenger that communicates information about adipose tissue energy stores to the central nervous system. Neuropeptide Y-containing neurons in the arcuate nucleus have been shown to express leptin receptors and, indeed, leptin has been shown to decrease neuropeptide Y mRNA levels in vivo and to decrease neuropeptide Y release from hypothalamic slices in vitro (Stephens et al., 1995). Thus, neuropeptide Y is probably one of the downstream mediators of leptin action in the brain. This conclusion is supported by the observation that the obese phenotype of the leptin-deficient ob/ob mouse is partially ameliorated by genetic deletion of neuropeptide Y (see above; Erickson et al., 1996b).

Activation of the melanocortin MC₃ and MC₄ receptors by melanocortin peptides such as α -melanocyte-stimulating hormone (α-MSH) decreases food intake, increases energy expenditure and decreases body weight (Vergoni and Bertolini, 2000). Conversely, blockade of these receptors by an endogenous antagonist known as agouti-related peptide (AGRP) has the opposite effect (Dinulescu and Cone, 2000). Furthermore, genetic ablation of the melanocortin MC₃ or MC₄ receptor has been shown to cause obesity in both mice and, in the case of the melanocortin MC₄ receptor, in humans (Spiegelman and Flier, 2001). Interestingly, AGRP is expressed exclusively in neuropeptide Ycontaining neurons originating in the arcuate nucleus (Hahn et al., 1998). Thus, activation of arcuate neuropeptide Y neurons releases neuropeptide Y and AGRP, both of which increase food intake and body weight via activation of two distinct pathways. The obvious implication of this finding is that both arcuate neuropeptide Y/AGRP-containing neurons and arcuate melanocortin-containing neurons project to one or more common target neurons that express both neuropeptide Y receptors (Y₁ and/or Y₅; see below) and melanocortin receptors (MC₃ and/or MC₄). Since most arcuate melanocortin-containing neurons also express cocaine and amphetamine-regulated transcript (CART), another peptide that has been implicated in the central regulation of energy

homeostasis (Elias et al., 1998), these target neurons probably also express CART receptors. Identification of the neurotransmitters employed by these target neurons may unveil a "final common pathway" in the regulation of energy homeostasis and would thus suggest very interesting additional basic research and drug development approaches in the field of obesity. It has been suggested that these target neurons contain γ -aminobutyric acid (GABA), CART, thyrotropin-releasing hormone (TRH), and/or melanin concentrating hormone (MCH), but more work is required to confirm this (Spiegelman and Flier, 2001).

Neuropeptide Y has also been shown to interact with a number of other hormones, neurotransmitters and neuropeptides that are thought to play a role in the regulation of body weight (e.g., insulin, galanin, GABA, corticotropin-releasing hormone, melanin concentrating hormone, orexin, ghrelin, biogenic amines, opiate peptides) (Kalra et al., 1999; Spiegelman and Flier, 2001). Neuropeptide Y may also be involved in metabolic control of energy homeostasis as evidenced by the decrease in neuropeptide Y levels and the decrease in food intake seen after inhibition of fatty acid synthase (Loftus et al., 2000). Taken together, these data strongly support a role for neuropeptide Y in the complex and highly integrated brain neurochemical system that regulates energy homeostasis.

3.4. Role of neuropeptide Y receptor subtypes in the regulation of energy homeostasis

To design potential anti-obesity drugs, it is necessary to identify the neuropeptide Y receptor subtypes that mediate the ability of neuropeptide Y to promote positive energy balance and increased body weight. Several lines of experimental evidence now point to a primary role of the neuropeptide Y Y₁ and Y₅ receptors in mediating the effects of neuropeptide Y on energy homeostasis. Central administration of selective neuropeptide Y Y₁ or Y₅ receptor peptide agonists to rodents increases food intake, suggesting that activation of either neuropeptide Y receptor subtype results in an orexigenic response (Cabrele et al., 2000; Gerald et al., 1996; Mullins et al., 2001; Parker et al., 2000). Conversely, administration of selective neuropeptide Y Y₁ and Y₅ receptor antagonists partially decreases spontaneous, neuropeptide Y-induced and/or fasting-induced food intake (see below). Administration of neuropeptide Y Y₅ receptor antisense oligonucleotides also results in a decrease in food intake (Schaffhauser et al., 1997; Tang-Christensen et al., 1998). Furthermore, the pharmacology of the neuropeptide Y receptor mediating neuropeptide Y-induced feeding and decreases in body temperature and energy expenditure in rats strongly resembles that of the neuropeptide Y Y₅ receptor (Hwa et al., 1999). Both the neuropeptide Y Y₁ and Y₅ receptors are also regulated by changes in nutritional status (Widdowson et al., 1997b; Zammaretti et al., 2001). The neuropeptide Y Y₁ and Y₅ receptors have also been proposed to play a role in the central and peripheral effects of neuropeptide Y on plasma glucose and insulin levels (Bischoff and Michel, 1999; Burcelin et al., 2001).

Studies of mice lacking neuropeptide Y Y_1 receptors also support a role for this receptor in mediating the effects of neuropeptide Y on energy homeostasis. Neuropeptide Y Y_1 receptor-deficient mice exhibit slightly diminished natural and neuropeptide Y-stimulated feeding. In addition, feeding following a 24-h fast was significantly diminished by about 45% (Kushi et al., 1998; Pedrazzini et al., 1998). Paradoxically, neuropeptide Y Y_1 receptor-deficient mice develop mild late onset obesity and moderate hyperinsulinemia. These mice exhibit reduced locomotor activity during the nocturnal period, which could be one mechanism by which the late onset obesity occurs.

Studies of mice lacking neuropeptide Y Y₅ receptors have failed to provide strong support for a role of this receptor in mediating the effects of neuropeptide Y on energy homeostasis (Marsh et al., 1998). Neuropeptide Y Y₅ receptordeficient mice have normal growth and feeding when young. Core body temperature in neuropeptide Y Y₅ receptordeficient mice is normal, indicating a lack of a gross effect on metabolic rate. Feeding induced by low doses of neuropeptide Y was unchanged, while feeding induced by higher doses of neuropeptide Y was reduced in neuropeptide Y Y₅ receptor-deficient mice. Interestingly, neuropeptide Yinduced feeding was completely abolished in neuropeptide Y Y₅ receptor-deficient mice in which neuropeptide Y Y₁ receptors were also blocked by a neuropeptide Y Y₁ receptor antagonist (Marsh et al., 1998). This confirms that activation of both neuropeptide Y Y₁ and Y₅ receptors are responsible for neuropeptide Y-induced feeding. Neuropeptide Y Y₅deficient ob/ob mice were equally as obese as ob/ob mice, indicating that the attenuation of obesity in the ob/ob mouse lacking neuropeptide Y is not mediated through the Y₅ receptor (Marsh et al., 1998). Like the neuropeptide Y Y₁ receptor-deficient mice, neuropeptide Y Y₅ receptor-deficient mice unexpectedly develop a mild late onset obesity; however, this is more likely due to an increase in food intake.

The data from the neuropeptide Y Y₁ and Y₅ receptordeficient mice confirm that both receptors mediate neuropeptide Y-induced food intake and that the neuropeptide Y Y₁ receptor may also play a minor role in spontaneous food intake and a more significant role in food intake after deprivation. As discussed above, it is possible that these data underestimate the role of the neuropeptide Y Y_1 and Y_5 receptors in normal energy homeostasis if mice can readily compensate for their absence. For example, an increased involvement of the neuropeptide Y y₆ receptor in energy homeostasis may occur in the absence of the neuropeptide Y Y₁ or Y₅ receptor in mice. It is also possible that the neuropeptide Y Y₁ and Y₅ receptors are only involved in energy homeostasis under specific conditions (e.g., situations of negative energy balance such as starvation, obese patients who have lost substantial weight, etc.) It would also be interesting to examine energy homeostasis in mice lacking both the neuropeptide Y Y1 and Y5 receptors. Unfortunately, the close proximity of the genes for the neuropeptide $Y Y_1$ and Y_5 receptors precludes the generation of mice lacking both receptors via simple cross breeding of the individual knockout mice.

The selective neuropeptide Y Y2 receptor agonist C2neuropeptide Y does not increase food intake, suggesting that the receptor does not mediate the effects of neuropeptide Y on energy homeostasis (Gerald et al., 1996). However, mice deficient for the neuropeptide Y Y2 receptor subtype have increased body weight, food intake and fat deposition. Regulation of feeding and body weight is normal upon refeeding after starvation. Neuropeptide Y Y₂ receptor-deficient mice maintain the feeding response elicited by neuropeptide Y, but have an attenuated response to leptin administration (Naveilhan et al., 1999). These data suggest a possible role for the neuropeptide Y Y₂ receptor in normal energy homeostasis and in the response to leptin. The effect of the neuropeptide Y Y2 receptor may be indirect, due to effects of neuropeptide Y Y2 receptor activation on the release of neuropeptide Y and other neurotransmitters (see above). The recent disclosure of a non-peptide neuropeptide Y Y₂ receptor antagonist, BIIE0246 (Doods et al., 1999), may provide a tool for further testing the role of the neuropeptide Y Y₂ receptor in the regulation of energy homeostasis.

The selective neuropeptide Y Y₄ receptor agonist rat pancreatic polypeptide does not increase food intake, suggesting that activation of the neuropeptide Y Y₄ receptor is not responsible for mediating the effects of neuropeptide Y on energy homeostasis (Gerald et al., 1996). Herzog et al. (2001) have recently generated neuropeptide Y Y₄ receptor knockout mice. These mice tend to be leaner than their wild type counterparts, although there was little or no difference in food intake, adipose tissue mass or serum levels of insulin, glucose, leptin and corticosterone. Interestingly, ob/ob mice lacking the neuropeptide Y Y₄ receptor had similar body weight, increased food intake, less fat mass, decreased liver weight, decreased liver lipid content and improved fertility relative to control ob/ob mice. These data may suggest a role for the neuropeptide Y Y₄ receptor in energy homeostasis, but further studies are required, particularly since obesity or leanness are proving to be common phenotypes of many knockout mice.

Overall, the data suggest a primary, direct role for the neuropeptide Y Y_1 and Y_5 receptors in mediating the effects of neuropeptide Y on energy homeostasis. The neuropeptide Y Y_2 and/or Y_4 receptors may play a subtle, perhaps indirect role in energy homeostasis, but further work is required to confirm this.

4. Development of non-peptide neuropeptide Y receptor antagonists as therapeutic agents for the treatment of obesity

Within the pharmaceutical industry, there has been great interest in the development of neuropeptide Y Y_1 and Y_5

receptor antagonists as potential drugs for obesity management. There is now a significant body of primary and patent literature describing potent and selective neuropeptide Y Y_1 receptor antagonists, and an even larger body of literature devoted to neuropeptide Y Y_5 receptor antagonists. The following discussion focuses on the pharmacology of nonpeptidic neuropeptide Y receptor antagonists, with emphasis on reports from the primary literature that attempt to correlate neuropeptide Y receptor subtype-selective antagonism with effects on food intake and body weight. Several excellent reviews provide a comprehensive discussion of neuropeptide Y receptor antagonists that have been disclosed (Carpino, 2000; Hammond, 2001; Ling, 1999; Wieland et al., 2000; Zimanyi and Poindexter, 2000).

4.1. Neuropeptide $Y Y_1$ receptor antagonists

The discovery of potent and selective neuropeptide Y Y_1 receptor antagonists with oral bioavailability and central nervous system (CNS) penetrability has proved to be a challenging endeavor. Initially reported neuropeptide Y Y_1 receptor antagonists failed to cross the blood-brain barrier, and only recently have orally available brain penetrant neuropeptide Y Y_1 receptor antagonists been developed.

The first potent and selective non-peptidic neuropeptide Y Y₁ receptor antagonist, BIBP3226 (Rudolf et al., 1994), was designed to mimic the C-terminal region of neuropeptide Y (Fig. 1; Grundemar and Bloom, 1997). Structure—activity relationship studies confirmed that correct spatial arrangement with respect to the guanidine, phenol, and diphenylacetamide are critical for nanomolar affinity of

BIBP3226 at the neuropeptide Y Y_1 receptor (Grundemar and Bloom, 1997). Initially, it was shown that BIBP3226 has high affinity for human and rat neuropeptide Y Y_1 receptors (human Y_1 K_i =5.1 nM, rat Y_1 K_i =6.8 nM) and is inactive at neuropeptide Y Y_2 receptors (K_i >1000 nM) (Rudolf et al., 1994; Wieland et al., 1995). It was subsequently shown that BIBP3226 does not have significant affinity for neuropeptide Y Y_4 and Y_5 receptors (Gerald et al., 1996). BIBP3226 is perhaps the most widely studied neuropeptide Y receptor antagonist, and its use as a tool to investigate the vascular pharmacology of neuropeptide Y mediated by the neuropeptide Y Y_1 receptor has been reported (Doods et al., 1996).

Since BIBP3226 does not penetrate the blood-brain barrier (Doods et al., 1996), central administration of the compound is necessary to study the role of CNS neuropeptide Y Y₁ receptors in the regulation of food intake and energy expenditure. Administration of BIBP3226 intracere-broventricularly (Kask et al., 1998; O'Shea et al., 1997) or into the paraventricular nucleus (Morgan et al., 1998) was reported to block neuropeptide Y-induced food intake. However, observations consistent with CNS toxicity were seen on administration of BIBP3226 by these routes and these observations have cast doubt on the specificity of these effects (Doods et al., 1996; Morgan et al., 1998).

A structurally related analogue, BIBO3304, is claimed to cause less CNS toxicity than BIBP3226 (Fig. 1; Wieland et al., 1998). This compound has high affinity for cloned rat and human neuropeptide Y Y₁ receptors (rat Y₁ IC₅₀=0.72 nM, human IC₅₀=0.38 nM), and no significant affinity for the other neuropeptide Y receptor subtypes (IC₅₀>10.000 nM). In cells expressing the neuropeptide Y Y₁ receptor,

Fig. 1. Structures of neuropeptide Y Y₁ receptor antagonists.

BIBO3304 blocked neuropeptide Y-induced inhibition of forskolin-stimulated cAMP synthesis (p K_b =9.1), confirming that the compound is an antagonist. Central administration of BIBO3304 to rats attenuated feeding induced by neuropeptide Y and by food deprivation. There was no effect of BIBO3304 on feeding induced by noradrenaline or galanin and the inactive enantiomer of BIBO3304 (Y₁ K_i>1000 nM) did not inhibit neuropeptide Y-elicited food intake or feeding after food deprivation. These data suggest that the effects of BIBO3304 on food intake were mediated specifically by blockade of the neuropeptide Y Y₁ receptor (Wieland et al., 1998). In addition, intracerebroventricular administration of BIBO3304 to Zucker and Wistar rats for 1 week decreased body weight gain (Doods et al., 1997). These data indicate that activation of the neuropeptide Y Y₁ receptor promotes food intake and body weight gain under physiological and pathophysiological conditions, and that neuropeptide Y Y₁ receptor antagonists may be useful anti-obesity agents.

A group from Eli Lilly reported the first neuropeptide Y Y_1 receptor antagonist with sub-nanomolar potency, the substituted indole derivative LY-357897 (Fig. 1; Hipskind et al., 1997). The discovery of LY-357897 was achieved by a classical medicinal chemistry approach from a screening lead with weak affinity for the neuropeptide Y Y_1 receptor. An important conceptual step in the discovery of LY-357897 was appendage of a basic group to the indole nitrogen to improve potency. This strategy was adopted due to the importance of the dibasic functionality of the C-terminal region of neuropeptide Y for neuropeptide Y binding to the neuropeptide Y Y_1 receptor.

LY-357897 displayed a K_i of 0.75 nM at cloned human neuropeptide Y Y₁ receptors, and no affinity at cloned human neuropeptide Y Y_2 , Y_4 , and Y_5 receptors ($K_i > 10$ μM). The compound antagonized neuropeptide Y-induced Ca²⁺ mobilization (IC₅₀=3.2 nM) and neuropeptide Yinduced inhibition of forskolin-stimulated cAMP synthesis in cells expressing the human neuropeptide Y Y₁ receptor (IC₅₀=1.8 nM). Unfortunately, the compound could not be evaluated systemically due to a lack of oral and subcutaneous bioavailability. However, LY-357897 administered into the lateral ventricle of mice dose-dependently blocked food consumption elicited by intracerebroventricularly administered neuropeptide Y (ED₅₀=17 nmol). It was reported that at doses of LY-357897 that blocked neuropeptide Y-induced food intake, no changes in motor function or behavioral effects were observed, consistent with a neuropeptide Y Y1 receptor-specific mechanism for the effect.

Banyu workers have reported studies with the potent and selective neuropeptide Y Y_1 receptor antagonists J-104870 and J-115814 from a diaminopyridine series (Fig. 1; Kanatani et al., 1999, 2001). J-104870 is the first reported potent, orally bioavailable, brain penetrant neuropeptide Y Y_1 receptor antagonist (Kanatani et al., 1999). The compound has high affinity for cloned rat and human neuropeptide Y Y_1 receptors (human Y_1 K_i =0.26 nM, rat Y_1 K_i =0.51 nM) and no significant affinity for human neuropeptide Y Y_2 ,

 Y_4 , and Y_5 receptors ($K_1 > 1000$ nM). J-104870 dose-dependently inhibited the neuropeptide Y-induced increase in intracellular Ca²⁺ levels in cells expressing human neuropeptide Y Y₁ receptors (IC₅₀=3.2 nM), confirming that it is an antagonist. Oral administration of J-104870 (100 mg/kg) to Zucker rats suppressed spontaneous feeding over 24 h by 18%, affording brain levels of 0.5 μM 24 h after dosing. A similar reduction in spontaneous food intake was observed on intracerebroventricular injection of J-104870 to Zucker rats. In contrast, J-104870 had no effect on spontaneous food intake when administered intracerebroventricularly to satiated Sprague–Dawley rats, although the compound did block neuropeptide Y-induced food intake. These data support the conclusion that elevated neuropeptide Y levels present in the brains of Zucker rats are partially responsible for the obese phenotype of these rats.

The related compound J-115814 showed high affinity for human, rat and mouse neuropeptide Y Y₁ receptors (human $Y_1 K_i = 1.4 \text{ nM}$; rat $Y_1 K_i = 1.8 \text{ nM}$; mouse $Y_1 K_i = 1.9 \text{ nM}$). Much lower affinity was seen at human neuropeptide Y Y₂ $(K_i > 10,000 \text{ nM}), Y_4 (K_i = 620 \text{ nM}) \text{ and } Y_5 (K_i = 6000 \text{ nM})$ receptors, as well as mouse neuropeptide Y y6 receptors $(K_i > 10,000 \text{ nM})$ (Kanatani et al., 2001). Furthermore, the compound had no significant activity at 50 other receptors tested. The antagonist character of J-115814 was confirmed by its inhibition of neuropeptide Y-induced increases in intracellular Ca2+ levels in cells expressing human neuropeptide Y Y₁ receptors (IC₅₀=6.8 nM). Administration of J-115814 intravenously or intracerebroventricularly to satiated rats dose responsively suppressed neuropeptide Y-induced food intake by up to 50%. There was no effect on food intake, nor were any behavioral changes observed when J-115814 was administered alone.

When J-115814 was administered intraperitoneally to wild type or neuropeptide Y Y₅ receptor-deficient mice, neuropeptide Y-induced food intake was inhibited by about 50%. However, J-115814 failed to suppress neuropeptide Yelicited food intake in Y₁ receptor-deficient mice, supporting the notion that J-115814 inhibits neuropeptide Y-induced food intake by a neuropeptide Y Y1 receptor-specific mechanism. When administered intraperitoneally to db/db mice or to C57BL6 mice, J-115814 caused a reduction in spontaneous feeding with a minimum effective dose of 10 mg/kg. Hypothalamic levels of J-115814 were consistent with the effects on food intake being mediated by neuropeptide Y Y₁ receptor blockade. The greater inhibition of food intake seen in db/db mice at the minimum effective dose is consistent with the proposal that increased activation of neuropeptide Y Y₁ receptors plays an important role in pathophysiological hyperphagia and obesity in db/db mice (Kanatani et al., 2001).

Recently, a group from Pfizer and Neurogen has also reported an orally bioavailable neuropeptide Y Y₁ receptor antagonist that crosses the blood-brain barrier, CP-671,906 (Fig. 1; Griffith et al., 2001). The compound showed high affinity for cloned rat and human neuropeptide Y Y₁ recep-

tors (rat Y_1 $K_i=3.5$ nM, human Y_1 $K_i=1.5$ nM), with no significant affinity for neuropeptide Y Y₂ and Y₅ receptors (K_i>1000 nM). In addition, CP-671,906 did not show significant activity at more than 35 other receptors, enzymes and ion channels. The functional antagonist activity of CP-671,906 was demonstrated by its inhibition of neuropeptide Y-induced intracellular Ca²⁺ flux in cells expressing neuropeptide Y Y₁ receptors ($K_b=11$ nM), as well as by its inhibition of neuropeptide Y-induced contractions of rabbit vas deferens ($K_b=26$ nM). In rats administered 40 mg/kg orally, CP-671,906 achieved plasma levels >1 μM and a brain to plasma ratio of 1. The compound blocked neuropeptide Y-induced food intake in rats when dosed 4 h prior to neuropeptide Y administration, which corresponded to the time at which brain interstitial fluid concentrations peaked at levels 10- to 70-fold over the K_i . Food intake elicited by the neuropeptide Y Y₅ receptor agonist bovine pancreatic polypeptide was not affected by CP-671,906, consistent with a neuropeptide Y Y₁ receptor-specific effect of the compound. CP-671,906 (40 mg/kg orally) inhibited refeeding in fooddeprived Wistar rats at 2, 4, and 24 h after dosing, but did not affect dark-cycle food intake in free-feeding rats. CP-671,906 (10 mg/kg orally) also decreased food intake in Zucker rats by 15% over 2 days, but caused adverse effects at

slightly higher doses (40 mg/kg orally). The compound also inhibited food intake in overweight beagle dogs, but behavioral changes were noted at the dose tested (10 mg/kg orally). Overall, it is difficult to rule out the possibility that the anorectic effects of CP-671,906 are due to a non-specific mechanism given that anorectic doses approach doses that produce adverse effects in several species.

In general, data with neuropeptide Y Y_1 receptor antagonists support a role for the neuropeptide Y Y_1 receptor in the modulation of energy homeostasis, particularly feeding induced by deprivation in normal rodents or feeding in genetically obese rodents. Orally active, brain penetrant neuropeptide Y Y_1 receptor antagonists have only recently been developed, but the compounds reported thus far are not very potent in vivo. It will likely be necessary to develop compounds with greater in vivo potency and/or a lower incidence of adverse effects.

4.2. Neuropeptide Y Y₅ receptor antagonists

The first potent, selective neuropeptide Y Y₅ receptor antagonist was the aminoquinazoline CGP 71683A reported in 1998 by a group from Novartis and Synaptic (Fig. 2; Criscione et al., 1998). Starting from a lead with weak

$$\begin{array}{c} NH_2 \\ NH$$

Fig. 2. Structures of neuropeptide Y Y₅ receptor antagonists.

affinity for neuropeptide Y Y₁ and Y₅ receptors, CGP 71683A was discovered through a process of combinatorial chemistry and traditional medicinal chemistry. Structure—activity relationship studies suggest that important pharmacophore elements of CGP 71683A are the hydrogen bond donor—acceptor properties of the aminoquinazoline ring and the hydrogen bond accepting sulfonamide separated by an appropriate distance by the hydrophobic spacer (Rueeger et al., 2000).

CGP 71683A has high affinity for the rat and human neuropeptide Y Y₅ receptors (rat Y₅ IC₅₀=1.4 nM; human Y₅ IC₅₀=2.9 nM) and low affinity for human neuropeptide Y Y₁, Y₂ and Y₄ receptors (IC₅₀>1000 nM). The compound was an antagonist of the rat neuropeptide Y Y₅ receptor, selectively inhibiting neuropeptide Y-induced intracellular Ca²⁺ transients in cells expressing neuropeptide Y Y₅ receptors. Intraperitoneal injection of CGP 71683A to satiated lean rats at a dose of 10 mg/kg inhibited neuropeptide Y-induced food intake by 50% and attenuated feeding after food deprivation, but had little effect on food intake in free-feeding rats. Rats that were chronically dosed with CGP 71683A showed reduced food intake and body weight gain over the first few days of treatment, but food intake and body weight gain subsequently approached control levels. A combination of intraperitoneally administered CGP 71683A and the neuropeptide Y Y₁ receptor antagonist BIBO3304 was recently reported to produce anorectic effects in lean rats, Zucker rats, and ob/ob mice at doses where either agent alone was ineffective (Duhault et al., 2000). CGP 71683A has recently been reported to have potent affinity for the 5-hydroxytryptamine (5-HT) reuptake recognition site and muscarinic receptors, so the possibility that CGP 71683A produces anorectic effects through mechanisms other than neuropeptide Y Y₅ receptor blockade cannot be ruled out (Della Zuana et al., 2001). Indeed, it was recently reported that the anorectic effects of CGP 71683A are identical in wild type and Y₅ receptor-deficient mice, thus indicating that the anorectic effects of the compound are non-specific (Criscione et al., 2001).

A novel series of neuropeptide Y Y₅ receptor antagonists based on a β-aminotetralin scaffold has been reported by a group at R.W. Johnson (Youngman et al., 2000). Potent analogues such as 1 and 2 were identified through optimization of a micromolar screening lead (Fig. 2). The latter compound contains the bis-(aminomethyl)cyclohexyl linker present in CGP71683A. Compounds 1 and 2 bind with high affinity to human neuropeptide Y Y₅ receptors (IC₅₀=23 and 1.0 nM, respectively), and inhibited peptide YY-induced [35S]GTPyS binding to membranes from Bowes melanoma cells transfected with neuropeptide Y Y₅ receptors. Furthermore, 1 showed no significant affinity for human neuropeptide Y Y₁ and Y₂ receptors, as well as for over 30 Gprotein coupled receptors and ion channels. Intraperitoneal administration of 1 (30 mg/kg) to fasted rats was reported to significantly reduce food consumption by 35% 6 h after dosing. While compounds in the series were said to be generally well-tolerated, no data addressing the specificity of the effects of 1 on food intake or correlating CNS exposure to drug with the anorectic effects were reported in this study.

The R.W. Johnson group has also described a structurally distinct pyrazole series represented by 3 and 4 (Fig. 2; Kordik et al., 2001a,b). Compounds 3 and 4 were reported to show IC $_{50}$ s of 15 and 80 nM, respectively, against the human neuropeptide Y Y $_{5}$ receptor, and did not bind significantly to human neuropeptide Y Y $_{1}$ or Y $_{2}$ receptors. No data confirming the antagonistic properties of these compounds at neuropeptide Y Y $_{5}$ receptors was reported. At a dose of 30 mg/kg administered intraperitoneally to rats, 4 decreased fasting-induced feeding by 43% in the period 2–6 h after administration. Compound 4 was said to be well tolerated. However, in the absence of data to support neuropeptide Y Y $_{5}$ receptor specificity, the mechanism of the anorectic effects of 4, like 1, must be considered circumstantial and open to question.

L-152,804 is an orally bioavailable, brain-penetrant neuropeptide Y Y₅ receptor antagonist that has reasonable affinity for human and rat neuropeptide Y Y₅ receptors (human $Y_5 K_i = 26$ nM; rat $Y_5 K_i = 31$ nM) and low affinity for human neuropeptide Y Y1, Y2, and Y4 receptors (K_i>10,000 nM) (Fig. 2; Kanatani et al., 2000). The antagonist activity of L-152,804 was confirmed in cells expressing the human neuropeptide Y Y₅ receptor, in which L-152,804 inhibited the neuropeptide Y-induced increase in intracellular Ca²⁺ levels (IC₅₀=210 nM). When administered intracerebroventricularly to satiated rats, L-152,804 inhibited food intake elicited by the neuropeptide Y Y₄/Y₅ receptor-selective agonist, bovine pancreatic polypeptide. Administration of L-152,804 alone had no effect on food intake, and was reported not to induce overt behavioral changes, indicating that the anorectic effect of the compound is neuropeptide Y Y₅ receptor-specific. When administered orally to rats, L-152,804 (10 mg/kg) also inhibited food intake elicited by intracerebroventricular bovine pancreatic polypeptide. However, L-152,804 administered intracerebroventricularly or orally failed to inhibit neuropeptide Y-stimulated food intake, despite brain levels of 2.9 µM 2 h after oral administration. Based on these observations, it was concluded that activation of the neuropeptide Y₅ receptor does not substantially contribute to food intake in rodents. However, further studies with functionally more potent neuropeptide Y Y₅ receptor antagonists would address this more definitively.

A group from Pfizer and Neurogen has recently reported extensive pharmacological evaluation of a more potent, orally bioavailable, brain penetrant neuropeptide Y Y_5 receptor antagonist, the 2,4-diarylimidazole 5, in models of feeding and energy expenditure (Fig. 2; Elliott et al., 2001). Compound 5 showed high affinity for human neuropeptide Y Y_5 receptors (IC $_{50}$ =1.2 nM), and inhibited neuropeptide Y-induced Ca $^{2+}$ mobilization in Bowes melanoma cells expressing the neuropeptide Y Y_5 receptor (IC $_{50}$ =0.4 nM). The compound did not have significant affinity for human neuropeptide Y Y_1 or Y_2 receptors (IC $_{50}$ >1000 nM),

or for over 50 other receptors. Food intake elicited by bovine pancreatic polypeptide was inhibited by 56% after oral administration of compound 5 (30 mg/kg). Brain and CSF levels were found to be 4 and 0.2 μ M, respectively, 0.5 h after dosing. These data demonstrate that compound achieves excellent CNS exposure and can block a neuropeptide Y Y₅ receptor-specific effect in vivo. However, compound 5 at a dose of 40 mg/kg orally failed to inhibit feeding following food deprivation in rats, and had no effect on spontaneous feeding at a dose of 30 mg/kg orally. Thermogenic effects were not evident with compound 5, as it did not cause significant changes in oxygen consumption or respiratory quotient when dosed at 30 mg/kg orally. There was no effect of the compound on spontaneous locomotor activity, consistent with a lack of overt behavioral effects. These data indicate that while activation of the neuropeptide Y Y₅ receptor with an exogenous ligand can promote food intake, it does not play a significant role in regulation of food intake and energy expenditure in lean rats under physiological conditions.

4.3. Potential side effects of neuropeptide Y receptor antagonists

Because neuropeptide Y is involved in a wide variety of physiological processes, many of which are mediated via neuropeptide Y Y₁ and Y₅ receptors, it is possible that neuropeptide Y Y₁ and Y₅ receptor antagonists developed for the treatment of obesity will be associated with specific mechanism-based side effects. Neuropeptide Y Y₁ receptor activation is known to lead to increased blood pressure, proliferation of neuronal precursor cells, anxiolysis, analgesia, neurogenic inflammation and modulation of pituitary hormone secretion (Wahlestedt et al., 1993; Grundemar and Bloom, 1997; El Majdoubi et al., 2000; Hansel et al., 2001; Naveilhan et al., 2001). Interestingly, neuropeptide Y Y₁ receptor antagonists such as BIBP3226 have no effect on blood pressure, suggesting that hypotension will not be an issue with these drugs (Doods et al., 1996). There have been reports that neuropeptide Y Y₁ receptor antagonists are anxiogenic and accelerate the onset of puberty (Kask et al., 1996; Pralong et al., 2000). The latter effect presumably reflects effects of the neuropeptide Y Y₁ receptor antagonist on pituitary gonadotropin secretion. All studies reported to date used intracerebroventricular administration of neuropeptide Y Y₁ receptor antagonists and additional studies with newer, orally active, brain penetrant compounds would be more informative.

Similarly, neuropeptide Y Y_5 receptor activation has been associated with anti-epileptic effects, attenuation of opiate withdrawal, modulation of circadian rhythms, regulation of pituitary hormone secretion, natriuresis and decreases in plasma glucose (Bischoff and Michel, 1999; Marsh et al., 1999a; Raposinho et al., 1999; Yannielli and Harrington, 2001). Unfortunately, the effect of neuropeptide Y Y_5 receptor antagonists on these processes has not been reported.

5. Summary and perspective

Data from mice lacking neuropeptide Y or the neuropeptide Y₁ receptor suggest that activation of the neuropeptide Y Y₁ receptor by neuropeptide Y plays a role in maintaining normal food intake as well as food intake after deprivation. Studies with selective neuropeptide Y Y₁ receptor antagonists also suggest a role for neuropeptide Y Y₁ receptors in maintaining food intake under conditions of real or apparent deprivation (e.g., food-deprived animals and genetically obese animals in which leptin signaling to the hypothalamus has been disrupted). The role of the neuropeptide Y Y₅ receptor in energy homeostasis is less clear, however. Mice lacking the neuropeptide Y Y₅ receptor do not differ from wild type mice in paradigms that assess food intake and body weight under a variety of conditions. Furthermore, the reported effects of neuropeptide Y Y₅ receptor antagonists on food intake and body weight are conflicting. Although administration of some neuropeptide Y Y₅ receptor antagonists is associated with reduced food intake and body weight gain, evidence for the specificity of these effects is lacking.

The modest effects of neuropeptide Y deficiency, neuropeptide Y Y₁ receptor deficiency and neuropeptide Y Y₁ receptor antagonists and the lack of any consistent effect of neuropeptide Y Y₅ receptor deficiency and neuropeptide Y Y₅ receptor antagonists on energy homeostasis suggest that the role of neuropeptide Y in the regulation of body weight is more complicated than previously envisioned. Overall, these results may indicate that the high level of redundancy in the regulation of body weight insures that mice can substantially compensate for the loss of a single neuropeptide or neuropeptide receptor under normal conditions. However, the data obtained to date suggests that neuropeptide Y plays a critical role in energy homeostasis under very specific physiological conditions, particularly conditions of real or apparent deprivation. These data indicate that neuropeptide Y receptor antagonists may be most useful in human conditions where appetite is increased and energy expenditure is decreased due to activation of the starvation response. Such conditions include obese patients who are dieting, formerly obese patients who have lost substantial weight, and patients with complete or partial leptin deficiency.

Significant progress has been made in the identification of structurally diverse, orally bioavailable neuropeptide Y Y_1 and Y_5 receptor antagonists that cross the blood-brain barrier. However, there is a clear need for further studies with both neuropeptide Y Y_1 and Y_5 receptor antagonists in order to clarify their potential as anti-obesity agents. Most studies have been performed in lean rodents or in genetically obese rodents that do not mimic common obesity in the human population. It would be desirable to evaluate the effects of compounds in diet-induced obese rodents and non-rodents as disease models that more closely mimic human obesity. Other critical issues in the development of neuropeptide Y receptor antagonists as effective agents for

obesity management are the need to overcome counterbalancing effects of the multiple complementary mechanisms involved in energy homeostasis that tend to oppose any changes in body weight and the identification of patient subclasses most likely to benefit from treatment.

References

- Akabayashi, A., Wahlestedt, C., Alexander, J.T., Leibowitz, S.F., 1994.Specific inhibition of endogenous neuropeptide Y synthesis in arcuate nucleus by antisense oligonucleotides suppresses feeding behavior and insulin secretion. Brain Res. Mol. Brain Res. 21, 55–61.
- Bannon, A.W., Seda, J., Carmouche, M., Francis, J.M., Norman, M.H., Karbon, B., McCaleb, M.L., 2000. Behavioral characterization of neuropeptide Y knockout mice. Brain Res. 868, 79–87.
- Bergen, H.T., Mizuno, T., Taylor, J., Mobbs, C.V., 1999. Resistance to dietinduced obesity is associated with increased proopiomelanocortin mRNA and decreased neuropeptide Y mRNA in the hypothalamus. Brain Res. 851, 198–203.
- Bischoff, A., Michel, M.C., 1999. Emerging functions for neuropeptide Y5 receptors. Trends Pharmacol. Sci. 20, 104–106.
- Burcelin, R., Brunner, H., Seydoux, J., Thorensa, B., Pedrazzini, T., 2001. Increased insulin concentrations and glucose storage in neuropeptide Y Y1 receptor-deficient mice. Peptides 22, 421–427.
- Burkhoff, A., Linemeyer, D.L., Salon, J.A., 1998. Distribution of a novel hypothalamic neuropeptide Y receptor gene and its absence in rat. Brain Res. Mol. Brain Res. 53, 311–316.
- Caberlotto, L., Fuxe, K., Sedvall, G., Hurd, Y.L., 1997. Localization of neuropeptide Y Y1 mRNA in the human brain: abundant expression in cerebral cortex and striatum. Eur. J. Neurosci. 9, 1212–1225.
- Caberlotto, L., Fuxe, K., Rimland, J.M., Sedvall, G., Hurd, Y.L., 1998a. Regional distribution of neuropeptide Y Y2 receptor messenger RNA in the human post mortem brain. Neuroscience 86, 167–178.
- Caberlotto, L., Tinner, B., Bunnemann, B., Agnati, L., Fuxe, K., 1998b. On the relationship of neuropeptide Y Y1 receptor-immunoreactive neuronal structures to the neuropeptide Y-immunoreactive nerve terminal networks. A double immunolabelling analysis in the rat brain. Neuroscience 86, 827–845.
- Caberlotto, L., Fuxe, K., Hurd, Y.L., 2000. Characterization of NPY mRNA-expressing cells in the human brain: co-localization with Y2 but not Y1 mRNA in the cerebral cortex, hippocampus, amygdala, and striatum. J. Chem. Neuroanat. 20, 327–337.
- Cabrele, C., Langer, M., Bader, R., Wieland, H.A., Doods, H.N., Zerbe, O., Beck-Sickinger, A.G., 2000. The first selective agonist for the neuropeptide YY5 receptor increases food intake in rats. J. Biol. Chem. 275, 36043-36048.
- Carpino, P.A., 2000. Patent focus on new obesity agents: September 1999– February 2000. Exp. Opin. Ther. Patents 10, 819–831.
- Cerda-Reverter, J.M., Larhammar, D., 2000. Neuropeptide Y family of peptides: structure, anatomical expression, function, and molecular evolution. Biochem. Cell Biol. 78, 371–392.
- Chronwall, B.M., DiMaggio, D.A., Massari, V.J., Pickel, V.M., Ruggiero, D.A., O'Donohue, T.L., 1985. The anatomy of neuropeptide-Y-containing neurons in rat brain. Neuroscience 15, 1159–1181.
- Clark, J.T., Kalra, P.S., Crowley, W.R., Kalra, S.P., 1984. Neuropeptide Y and human pancreatic polypeptide stimulate feeding behavior in rats. Endocrinology 115, 427–429.
- Criscione, L., Rigollier, P., Batzl-Hartmann, C., Rueger, H., Stricker-Krongrad, A., Wyss, P., Brunner, L., Whitebread, S., Yamaguchi, Y., Gerald, C., Heurich, R.O., Walker, M.W., Chiesi, M., Schilling, W., Hofbauer, K.G., Levens, N., 1998. Food intake in free-feeding and energy-deprived lean rats is mediated by the neuropeptide Y5 receptor. J. Clin. Invest. 102, 2136–2145.
- Criscione, L., Schaffhauser, A.O., Guerini, D., Schmid, H., Buhlmayer, P.,

- 2001. Effects of the Y5 receptor antagonist CGP71683A on food intake in male Y5 receptor knockout mice. Sixth International NPY Conference, Sydney, Australia.
- Della Zuana, O., Sadlo, M., Germain, M., Feletou, M., Chamorro, S., Tisserand, F., Montrion, C., Boivin, J.F., Duhault, J., Boutin, J.A., Levens, N., 2001. Reduced food intake in response to CGP 71683A may be due to mechanisms other than NPY Y5 receptor blockade. Int. J. Obes. Relat. Metab. Disord. 25, 84–94.
- Dinulescu, D.M., Cone, R.D., 2000. Agouti and agouti-related protein: analogies and contrasts. J. Biol. Chem. 275, 6695–6698.
- Doods, H.N., Wieland, H.A., Engel, W., Eberlein, W., Willim, K.D., Entzeroth, M., Wienen, W., Rudolf, K., 1996. BIBP 3226, the first selective neuropeptide Y1 receptor antagonist: a review of its pharmacological properties. Regul. Pept. 65, 71–77.
- Doods, H.N., Engel, W., Wieland, H.A., Eberlein, W., Rudolf, K., Judge, M., Hamilton, B.S., 1997. Effects of the novel Y1 antagonist BI-BO3304 on feeding in rodents. Regul. Pept. 71, 212.
- Doods, H., Gaida, W., Wieland, H.A., Dollinger, H., Schnorrenberg, G., Esser, F., Engel, W., Eberlein, W., Rudolf, K., 1999. BIIE0246: a selective and high affinity neuropeptide Y Y(2) receptor antagonist. Eur. J. Pharmacol. 384, R3–R5.
- Duhault, J., Boulanger, M., Chamorro, S., Boutin, J.A., Della Zuana, O., Douillet, E., Fauchere, J.L., Feletou, M., Germain, M., Husson, B., Vega, A.M., Renard, P., Tisserand, F., 2000. Food intake regulation in rodents: Y5 or Y1 NPY receptors or both? Can. J. Physiol. Pharmacol. 78, 173–185.
- Dumont, Y., Fournier, A., St-Pierre, S., Quirion, R., 1996. Autoradiographic distribution of [1251]Leu31,Pro34]PYY and [1251]PYY3-36 binding sites in the rat brain evaluated with two newly developed Y1 and Y2 receptor radioligands. Synapse 22, 139–158.
- Dumont, Y., Fournier, A., Quirion, R., 1998. Expression and characterization of the neuropeptide Y Y5 receptor subtype in the rat brain. J. Neurosci. 18, 5565–5574.
- Egawa, M., Yoshimatsu, H., Bray, G.A., 1991. Neuropeptide Y suppresses sympathetic activity to interscapular brown adipose tissue in rats. Am. J. Physiol. 260, R328–R334.
- Elias, C.F., Lee, C., Kelly, J., Aschkenasi, C., Ahima, R.S., Couceyro, P.R., Kuhar, M.J., Saper, C.B., Elmquist, J.K., 1998. Leptin activates hypothalamic CART neurons projecting to the spinal cord. Neuron 21, 1375–1385.
- Elliott, R.L., Oliver, R.M., Hammond, M., Patterson, T.A., She, L., Hargrove, D.M., Martin, K.A., Maurer, T.S., Kalvass, J.C., Morgan, B.P., DaSilva-Jardine, P.A., Stevenson, R.W., Mack, C.M., Cassella, J.V., 2001. The in vitro and in vivo characterization of 3-{2-[6-(2-tert-butoxyethoxy)-pyridin-3-yl]-1H-imidazol-4-yl}-benzonitrile, (I), a potent and selective NPY-Y5 antagonist. Sixth International NPY Conference, Sydney, Australia.
- El Majdoubi, M., Sahu, A., Ramaswamy, S., Plant, T.M., 2000. Neuropeptide Y: a hypothalamic brake restraining the onset of puberty in primates. Proc. Natl. Acad. Sci. U. S. A. 97, 6179–6184.
- Erickson, J.C., Clegg, K.E., Palmiter, R.D., 1996a. Sensitivity to leptin and susceptibility to seizures of mice lacking neuropeptide Y. Nature 381, 415–418
- Erickson, J.C., Hollopeter, G., Palmiter, R.D., 1996b. Attenuation of the obesity syndrome of ob/ob mice by the loss of neuropeptide Y. Science 274, 1704–1707.
- Erickson, J.C., Ahima, R.S., Hollopeter, G., Flier, J.S., Palmiter, R.D., 1997. Endocrine function of neuropeptide Y knockout mice. Regul. Pept. 70, 199–202.
- Flynn, M.C., Turrin, N.P., Plata-Salaman, C.R., Ffrench-Mullen, J.M., 1999. Feeding response to neuropeptide Y-related compounds in rats treated with Y5 receptor antisense or sense phosphothio-oligodeoxynucleotide. Physiol. Behav. 66, 881–884.
- Gehlert, D.R., Beavers, L.S., Johnson, D., Gackenheimer, S.L., Schober, D.A., Gadski, R.A., 1996. Expression cloning of a human brain neuropeptide Y Y2 receptor. Mol. Pharmacol. 49, 224–228.
- Gerald, C., Walker, M.W., Criscione, L., Gustafson, E.L., Batzl-Hartmann,

- C., Smith, K.E., Vaysse, P., Durkin, M.M., Laz, T.M., Linemeyer, D.L., Schaffhauser, A.O., Whitebread, S., Hofbauer, K.G., Taber, R.I., Branchek, T.A., Weinshank, R.L., 1996. A receptor subtype involved in neuropeptide-Y-induced food intake. Nature 382, 168–171.
- Goumain, M., Voisin, T., Lorinet, A.M., Laburthe, M., 1998. Identification and distribution of mRNA encoding the Y1, Y2, Y4, and Y5 receptors for peptides of the PP-fold family in the rat intestine and colon. Biochem. Biophys. Res. Commun. 247, 52–56.
- Goumain, M., Voisin, T., Lorinet, A.M., Ducroc, R., Tsocas, A., Roze, C., Rouet-Benzineb, P., Herzog, H., Balasubramaniam, A., Laburthe, M., 2001. The peptide YY-preferring receptor mediating inhibition of small intestinal secretion is a peripheral Y(2) receptor: pharmacological evidence and molecular cloning. Mol. Pharmacol. 60, 124–134.
- Griffith, D.A., Blum, C.A., Carpino, P.A., Cassella, J., Darrow, J.W., De Lombaert, S., Hargrove, D.M., Hickman, M.A., Mack, C.M., Maurer, T.S., Sanders, M.J., Ashton, M.A., Giangiordano, M., He, P., Inthavongsay, J.K., Klade, L.E., Lebel, W.S., Martin, K.A., Regan, C., Rose, C.R., Tran, J., Vage, C., 2001. Structure—activity relationships within a series of pyrazolopyrimidine NPY Y1 receptor antagonists. 222nd ACS National Meeting, Chicago, IL.
- Grundemar, L., Bloom, S.R., 1997. Neuropeptide Y and Drug Development Academic Press, San Diego.
- Grundemar, L., Hakanson, R., 1993. Multiple neuropeptide Y receptors are involved in cardiovascular regulation. Peripheral and central mechanisms. Gen. Pharmacol. 24, 785–796.
- Guan, X.M., Yu, H., Trumbauer, M., Frazier, E., Van der Ploeg, L.H., Chen, H., 1998. Induction of neuropeptide Y expression in dorsomedial hypothalamus of diet-induced obese mice. NeuroReport 9, 3415–3419.
- Hahn, T.M., Breininger, J.F., Baskin, D.G., Schwartz, M.W., 1998. Coexpression of AGRP and NPY in fasting-activated hypothalamic neurons. Nat. Neurosci. 1, 271–272.
- Hammond, M., 2001. Neuropeptide Y receptor antagonists. IDrugs 4, 920–927.
- Hansel, D.E., Eipper, B.A., Ronnett, G.V., 2001. Neuropeptide Y functions as a neuroproliferative factor. Nature 410, 940-944.
- Herzog, H., Couzens, M., Ormandy, C., Sainsbury, A., 2001. Y4 receptor deletion improves fertility in ob/ob mice without affecting the obese phenotype. Sixth International NPY Conference, Sydney, Australia.
- Higuchi, H., Costa, E., Yang, H.Y., 1988. Neuropeptide Y inhibits the nicotine-mediated release of catecholamines from bovine adrenal chromaffin cells. J. Pharmacol. Exp. Ther. 244, 468–474.
- Hipskind, P.A., Lobb, K.L., Nixon, J.A., Britton, T.C., Bruns, R.F., Catlow, J., Dieckman-McGinty, D.K., Gackenheimer, S.L., Gitter, B.D., Iyengar, S., Schober, D.A., Simmons, R.M., Swanson, S., Zarrinmayeh, H., Zimmerman, D.M., Gehlert, D.R., 1997. Potent and selective 1,2,3-trisubstituted indole NPY Y-1 antagonists. J. Med. Chem. 40, 3712–3714.
- Hollopeter, G., Erickson, J.C., Palmiter, R.D., 1998. Role of neuropeptide Y in diet-, chemical- and genetic-induced obesity of mice. Int. J. Obes. Relat. Metab. Disord. 22, 506–512.
- Hwa, J.J., Witten, M.B., Williams, P., Ghibaudi, L., Gao, J., Salisbury, B.G., Mullins, D., Hamud, F., Strader, C.D., Parker, E.M., 1999. Activation of the NPY Y5 receptor regulates both feeding and energy expenditure. Am. J. Physiol. 277, R1428–R1434.
- Kaga, T., Inui, A., Okita, M., Asakawa, A., Ueno, N., Kasuga, M., Fujimiya, M., Nishimura, N., Dobashi, R., Morimoto, Y., Liu, I.M., Cheng, J.T., 2001. Modest overexpression of neuropeptide Y in the brain leads to obesity after high-sucrose feeding. Diabetes 50, 1206–1210.
- Kalra, S.P., Dube, M.G., Pu, S., Xu, B., Horvath, T.L., Kalra, P.S., 1999. Interacting appetite-regulating pathways in the hypothalamic regulation of body weight. Endocr. Rev. 20, 68–100.
- Kanatani, A., Kanno, T., Ishihara, A., Hata, M., Sakuraba, A., Tanaka, T., Tsuchiya, Y., Mase, T., Fukuroda, T., Fukami, T., Ihara, M., 1999. The novel neuropeptide Y Y(1) receptor antagonist J-104870: a potent feeding suppressant with oral bioavailability. Biochem. Biophys. Res. Commun. 266, 88–91.
- Kanatani, A., Ishihara, A., Iwaasa, H., Nakamura, K., Okamoto, O., Hidaka, M., Ito, J., Fukuroda, T., MacNeil, D., Van der Ploeg, L.H.T., Ishii,

- Y., Okabe, T., Fukami, T., Ihara, M., 2000. L-152,804: orally active and selective neuropeptide Y Y5 receptor antagonist. Biochem. Biophys. Res. Commun. 272, 169–173.
- Kanatani, A., Hata, M., Mashiko, S., Ishihara, A., Okamoto, O., Haga, Y., Ohe, T., Kanno, T., Murai, N., Ishii, Y., Fukuroda, T., Fukami, T., Ihara, M., 2001. A typical Y1 receptor regulates feeding behaviors: effects of a potent and selective Y1 antagonist, J-115814. Mol. Pharmacol. 59, 501–505.
- Kask, A., Rago, L., Harro, J., 1996. Anxiogenic-like effect of the neuropeptide Y Y1 receptor antagonist BIBP3226: antagonism with diazepam. Eur. J. Pharmacol. 317, R3-R4.
- Kask, A., Rago, L., Harro, J., 1998. Evidence for involvement of neuropeptide Y receptors in the regulation of food intake: studies with Y1selective antagonist BIBP3226. Br. J. Pharmacol. 124, 1507–1515.
- King, P.J., Williams, G., Doods, H., Widdowson, P.S., 2000. Effect of a selective neuropeptide Y Y(2) receptor antagonist, BIIE0246 on neuropeptide Y release. Eur. J. Pharmacol. 396, R1-R3.
- Kordik, C.P., Luo, C., Zanoni, B.C., Dax, S.L., McNally, J.J., Lovenberg, T.W., Wilson, S.J., Reitz, A.B., 2001a. Aminopyrazoles with high affinity for the human neuropeptide Y5 receptor. Bioorg. Med. Chem. Lett. 11, 2283–2286.
- Kordik, C.P., Luo, C., Zanoni, B.C., Lovenberg, T.W., Wilson, S.J., Vaidya, A.H., Crooke, J.J., Rosenthal, D.I., Reitz, A.B., 2001b. Pyrazolecarboxamide human neuropeptide Y5 receptor ligands with in vivo antifeedant activity. Bioorg. Med. Chem. Lett. 11, 2287–2290.
- Kushi, A., Sasai, H., Koizumi, H., Takeda, N., Yokoyama, M., Nakamura, M., 1998. Obesity and mild hyperinsulinemia found in neuropeptide Y-Y1 receptor-deficient mice. Proc. Natl. Acad. Sci. U. S. A. 95, 15659-15664
- Laburthe, M., Chenut, B., Rouyer-Fessard, C., Tatemoto, K., Couvineau, A., Servin, A., Amiranoff, B., 1986. Interaction of peptide YY with rat intestinal epithelial plasma membranes: binding of the radioiodinated peptide. Endocrinology 118, 1910–1917.
- Levin, B.E., Dunn-Meynell, A.A., 1997. Dysregulation of arcuate nucleus preproneuropeptide Y mRNA in diet-induced obese rats. Am. J. Physiol. 272. R1365—R1370.
- Ling, A.L., 1999. Neuropeptide Y receptor antagonists. Exp. Opin. Ther. Patents 9, 375–384.
- Loftus, T.M., Jaworsky, D.E., Frehywot, G.L., Townsend, C.A., Ronnett, G.V., Lane, M.D., Kuhajda, F.P., 2000. Reduced food intake and body weight in mice treated with fatty acid synthase inhibitors. Science 288, 2379–2381
- Lundberg, J.M., Terenius, L., Hokfelt, T., Martling, C.R., Tatemoto, K., Mutt, V., Polak, J., Bloom, S., Goldstein, M., 1982. Neuropeptide Y (NPY)-like immunoreactivity in peripheral noradrenergic neurons and effects of NPY on sympathetic function. Acta Physiol. Scand. 116, 477–480.
- Lundell, I., Blomqvist, A.G., Berglund, M.M., Schober, D.A., Johnson, D., Statnick, M.A., Gadski, R.A., Gehlert, D.R., Larhammar, D., 1995. Cloning of a human receptor of the NPY receptor family with high affinity for pancreatic polypeptide and peptide YY. J. Biol. Chem. 270, 29123–29128.
- Lundell, I., Statnick, M.A., Johnson, D., Schober, D.A., Starback, P., Gehlert, D.R., Larhammar, D., 1996. The cloned rat pancreatic polypeptide receptor exhibits profound differences to the orthologous receptor. Proc. Natl. Acad. Sci. U. S. A. 93, 5111–5115.
- Marks, J.L., Waite, K., 1996. Some acute effects of intracerebroventricular neuropeptide Y on insulin secretion and glucose metabolism in the rat. J. Neuroendocrinol. 8, 507–513.
- Marsh, D.J., Hollopeter, G., Kafer, K.E., Palmiter, R.D., 1998. Role of the Y5 neuropeptide Y receptor in feeding and obesity. Nat. Med. 4, 718–721.
- Marsh, D.J., Baraban, S.C., Hollopeter, G., Palmiter, R.D., 1999a. Role of the Y5 neuropeptide Y receptor in limbic seizures. Proc. Natl. Acad. Sci. U. S. A. 96, 13518–13523.
- Marsh, D.J., Miura, G.I., Yagaloff, K.A., Schwartz, M.W., Barsh, G.S., Palmiter, R.D., 1999b. Effects of neuropeptide Y deficiency on hypo-

- thalamic agouti-related protein expression and responsiveness to melanocortin analogues. Brain Res. 848, 66–77.
- McDermott, B.J., Millar, B.C., Piper, H.M., 1993. Cardiovascular effects of neuropeptide Y: receptor interactions and cellular mechanisms. Cardiovasc. Res. 27, 893–905.
- Michel, M.C., Beck-Sickinger, A., Cox, H., Doods, H.N., Herzog, H., Larhammar, D., Quirion, R., Schwartz, T., Westfall, T., 1998. XVI. International union of pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. Pharmacol. Rev. 50, 143–150.
- Morgan, D.G., Small, C.J., Abusnana, S., Turton, M., Gunn, I., Heath, M., Rossi, M., Goldstone, A.P., O'Shea, D., Meeran, K., Ghatei, M., Smith, D.M., Bloom, S., 1998. The NPY Y1 receptor antagonist BIBP 3226 blocks NPY induced feeding via a non-specific mechanism. Regul. Pept. 75–76, 377–382.
- Mullins, D.E., Guzzi, M., Xia, L., Parker, E.M., 2000. Pharmacological characterization of the cloned neuropeptide Y y₆ receptor. Eur. J. Pharmacol. 395, 87–93.
- Mullins, D., Kirby, D., Hwa, J., Guzzi, M., Rivier, J., Parker, E., 2001. Identification of potent and selective neuropeptide Y Y(1) receptor agonists with orexigenic activity in vivo. Mol. Pharmacol. 60, 534– 540.
- Nakamura, M., Sakanaka, C., Aoki, Y., Ogasawara, H., Tsuji, T., Kodama, H., Matsumoto, T., Shimizu, T., Noma, M., 1995. Identification of two isoforms of mouse neuropeptide Y-Y1 receptor generated by alternative splicing. Isolation, genomic structure, and functional expression of the receptors. J. Biol. Chem. 270, 30100-30102.
- Naveilhan, P., Neveu, I., Arenas, E., Ernfors, P., 1998. Complementary and overlapping expression of Y1, Y2 and Y5 receptors in the developing and adult mouse nervous system. Neuroscience 87, 289–302.
- Naveilhan, P., Hassani, H., Canals, J.M., Ekstrand, A.J., Larefalk, A., Chhajlani, V., Arenas, E., Gedda, K., Svensson, L., Thoren, P., Ernfors, P., 1999. Normal feeding behavior, body weight and leptin response require the neuropeptide Y Y2 receptor. Nat. Med. 5, 1188–1193.
- Naveilhan, P., Hassani, H., Lucas, G., Blakeman, K.H., Hao, J.X., Xu, X.J., Wiesenfeld-Hallin, Z., Thoren, P., Ernfors, P., 2001. Reduced antinociception and plasma extravasation in mice lacking a neuropeptide Y receptor. Nature 409, 513–517.
- Nichol, K.A., Morey, A., Couzens, M.H., Shine, J., Herzog, H., Cunningham, A.M., 1999. Conservation of expression of neuropeptide Y5 receptor between human and rat hypothalamus and limbic regions suggests an integral role in central neuroendocrine control. J. Neurosci. 19, 10295–10304.
- O'Shea, D., Morgan, D.G., Meeran, K., Edwards, C.M., Turton, M.D., Choi, S.J., Heath, M.M., Gunn, I., Taylor, G.M., Howard, J.K., Bloom, C.I., Small, C.J., Haddo, O., Ma, J.J., Callinan, W., Smith, D.M., Ghatei, M.A., Bloom, S.R., 1997. Neuropeptide Y induced feeding in the rat is mediated by a novel receptor. Endocrinology 138, 196–202.
- Parker, R.M., Herzog, H., 1999. Regional distribution of Y-receptor subtype mRNAs in rat brain. Eur. J. Neurosci. 11, 1431–1448.
- Parker, E.M., Balasubramaniam, A., Guzzi, M., Mullins, D.E., Salisbury, B.G., Sheriff, S., Witten, M.B., Hwa, J.J., 2000. [D-Trp(34)] neuropeptide Y is a potent and selective neuropeptide Y Y(5) receptor agonist with dramatic effects on food intake. Peptides 21, 393–399.
- Pedrazzini, T., Seydoux, J., Kunstner, P., Aubert, J.F., Grouzmann, E., Beermann, F., Brunner, H.R., 1998. Cardiovascular response, feeding behavior and locomotor activity in mice lacking the NPY Y1 receptor. Nat. Med. 4, 722–726.
- Pellieux, C., Sauthier, T., Domenighetti, A., Marsh, D.J., Palmiter, R.D., Brunner, H.R., Pedrazzini, T., 2000. Neuropeptide Y (NPY) potentiates phenylephrine-induced mitogen-activated protein kinase activation in primary cardiomyocytes via NPY Y5 receptors. Proc. Natl. Acad. Sci. U. S. A. 97, 1595–1600.
- Pralong, F.P., Voirol, M., Giacomini, M., Gaillard, R.C., Grouzmann, E.,
 2000. Acceleration of pubertal development following central blockade of the Y1 subtype of neuropeptide Y receptors. Regul. Pept. 95, 47–52.
 Raposinho, P.D., Broqua, P., Pierroz, D.D., Hayward, A., Dumont, Y.,

- Quirion, R., Junien, J.L., Aubert, M.L., 1999. Evidence that the inhibition of luteinizing hormone secretion exerted by central administration of neuropeptide Y (NPY) in the rat is predominantly mediated by the NPY-Y5 receptor subtype. Endocrinology 140, 4046–4055.
- Rudolf, K., Eberlein, W., Engel, W., Wieland, H.A., Willim, K.D., Entzeroth, M., Wienen, W., Beck-Sickinger, A.G., Doods, H.N., 1994. The first highly potent and selective non-peptide neuropeptide Y Y1 receptor antagonist: BIBP3226. Eur. J. Pharmacol. 271, R11–R13.
- Rueeger, H., Rigollier, P., Yamaguchi, Y., Schmidlin, T., Schilling, W., Criscione, L., Whitebread, S., Chiesi, M., Walker, M.W., Dhanoa, D., Islam, I., Zhang, J., Gluchowski, C., 2000. Design, synthesis and SAR of a series of 2-substituted 4-amino-quinazoline neuropeptide Y Y5 receptor antagonists. Bioorg. Med. Chem. Lett. 10, 1175–1179.
- Sahu, A., Kalra, P.S., Kalra, S.P., 1988. Food deprivation and ingestion induce reciprocal changes in neuropeptide Y concentrations in the paraventricular nucleus. Peptides 9, 83–86.
- Sahu, A., Sninsky, C.A., Kalra, P.S., Kalra, S.P., 1990. Neuropeptide-Y concentration in microdissected hypothalamic regions and in vitro release from the medial basal hypothalamus-preoptic area of streptozotocin-diabetic rats with and without insulin substitution therapy. Endocrinology 126, 192–198.
- Sainsbury, A., Cusin, I., Rohner-Jeanrenaud, F., Jeanrenaud, B., 1997. Adrenalectomy prevents the obesity syndrome produced by chronic central neuropeptide Y infusion in normal rats. Diabetes 46, 209–214.
- Sanacora, G., Kershaw, M., Finkelstein, J.A., White, J.D., 1990. Increased hypothalamic content of preproneuropeptide Y messenger ribonucleic acid in genetically obese Zucker rats and its regulation by food deprivation. Endocrinology 127, 730-737.
- Schaffhauser, A.O., Stricker-Krongrad, A., Brunner, L., Cumin, F., Gerald, C., Whitebread, S., Criscione, L., Hofbauer, K.G., 1997. Inhibition of food intake by neuropeptide Y Y5 receptor antisense oligodeoxynucleotides. Diabetes 46, 1792–1798.
- Shibasaki, T., Oda, T., Imaki, T., Ling, N., Demura, H., 1993. Injection of anti-neuropeptide Y gamma-globulin into the hypothalamic paraventricular nucleus decreases food intake in rats. Brain Res. 601, 313–316.
- Spiegelman, B.M., Flier, J.S., 2001. Obesity and the regulation of energy balance. Cell 104, 531–543.
- Stanley, B.G., Leibowitz, S.F., 1984. Neuropeptide Y: stimulation of feeding and drinking by injection into the paraventricular nucleus. Life Sci. 35, 2635–2642.
- Stanley, B.G., Magdalin, W., Seirafi, A., Thomas, W.J., Leibowitz, S.F., 1993. The perifornical area: the major focus of (a) patchily distributed hypothalamic neuropeptide Y-sensitive feeding system(s). Brain Res. 604, 304–317.
- Statnick, M.A., Schober, D.A., Gackenheimer, S., Johnson, D., Beavers, L., Mayne, N.G., Burnett, J.P., Gadski, R., Gehlert, D.R., 1998. Characterization of the neuropeptide Y5 receptor in the human hypothalamus: a lack of correlation between Y5 mRNA levels and binding sites. Brain Res. 810, 16–26.
- Stephens, T.W., Basinski, M., Bristow, P.K., Bue-Valleskey, J.M., Burgett, S.G., Craft, L., Hale, J., Hoffmann, J., Hsiung, H.M., Kriauciunas, A., MacKellar, W., Rosteck, P.R., Schoner, B., Smith, D., Tinsley, F.C., Zhang, X.-Y., Heiman, M., 1995. The role of neuropeptide Y in the antiobesity action of the obese gene product. Nature 377, 530–532.
- Tang-Christensen, M., Kristensen, P., Stidsen, C.E., Brand, C.L., Larsen, P.J., 1998. Central administration of Y5 receptor antisense decreases spontaneous food intake and attenuates feeding in response to exogenous neuropeptide Y. J. Endocrinol. 159, 307–312.
- Tatemoto, K., Carlquist, M., Mutt, V., 1982. Neuropeptide Y—a novel brain peptide with structural similarities to peptide YY and pancreatic polypeptide. Nature 296, 659–660.
- Vergoni, A.V., Bertolini, A., 2000. Role of melanocortins in the central control of feeding. Eur. J. Pharmacol. 405, 25-32.
- Vettor, R., Zarjevski, N., Cusin, I., Rohner-Jeanrenaud, F., Jeanrenaud, B., 1994. Induction and reversibility of an obesity syndrome by intracerebroventricular neuropeptide Y administration to normal rats. Diabetologia 37, 1202–1208.

- Wahlestedt, C., Pich, E.M., Koob, G.F., Yee, F., Heilig, M., 1993. Modulation of anxiety and neuropeptide Y-Y1 receptors by antisense oligodeoxynucleotides. Science 259, 528-531.
- Weinberg, D.H., Sirinathsinghji, D.J., Tan, C.P., Shiao, L.L., Morin, N., Rigby, M.R., Heavens, R.H., Rapoport, D.R., Bayne, M.L., Cascieri, M.A., Strader, C.D., Linemeyer, D.L., MacNeil, D.J., 1996. Cloning and expression of a novel neuropeptide Y receptor. J. Biol. Chem. 271, 16435–16438.
- Wettstein, J.G., Earley, B., Junien, J.L., 1995. Central nervous system pharmacology of neuropeptide Y. Pharmacol. Ther. 65, 397–414.
- Wharton, J., Gordon, L., Byrne, J., Herzog, H., Selbie, L.A., Moore, K., Sullivan, M.H., Elder, M.G., Moscoso, G., Taylor, K.M., Shine, J., Polak, J.M., 1993. Expression of the human neuropeptide tyrosine Y1 receptor. Proc. Natl. Acad. Sci. U. S. A. 90, 687–691.
- Widdowson, P.S., Buckingham, R., Williams, G., 1997a. Distribution of [Leu31,Pro34]NPY-sensitive, BIBP3226-insensitive [1251]PYY(3-36) binding sites in rat brain: possible relationship to Y5 NPY receptors. Brain Res. 778, 242-250.
- Widdowson, P.S., Upton, R., Henderson, L., Buckingham, R., Wilson, S., Williams, G., 1997b. Reciprocal regional changes in brain NPY receptor density during dietary restriction and dietary-induced obesity in the rat. Brain Res. 774, 1–10.
- Wieland, H.A., Willim, K.D., Entzeroth, M., Wienen, W., Rudolf, K., Eberlein, W., Engel, W., Doods, H.N., 1995. Subtype selectivity and antagonistic profile of the nonpeptide Y1 receptor antagonist BIBP 3226. J. Pharmacol. Exp. Ther. 275, 143–149.
- Wieland, H.A., Engel, W., Eberlein, W., Rudolf, K., Doods, H.N., 1998.Subtype selectivity of the novel nonpeptide neuropeptide Y Y1 receptor antagonist BIBO 3304 and its effect on feeding in rodents. Br. J. Pharmacol. 125, 549–555.

- Wieland, H.A., Hamilton, B.S., Krist, B., Doods, H.N., 2000. The role of NPY in metabolic homeostasis: implications for obesity therapy. Exp. Opin. Invest. Drugs 9, 1327–1346.
- Wilding, J.P., Gilbey, S.G., Mannan, M., Aslam, N., Ghatei, M.A., Bloom, S.R., 1992. Increased neuropeptide Y content in individual hypothalamic nuclei, but not neuropeptide Y mRNA, in diet-induced obesity in rats. J. Endocrinol. 132, 299–304.
- Yannielli, P.C., Harrington, M.E., 2001. The neuropeptide Y Y5 receptor mediates the blockade of "photic-like" NMDA-induced phase shifts in the golden hamster. J. Neurosci. 21, 5367–5373.
- Youngman, M.A., McNally, J.J., Lovenberg, T.W., Reitz, A.B., Willard, N.M., Nepomuceno, D.H., Wilson, S.J., Crooke, J.J., Rosenthal, D., Vaidya, A.H., Dax, S.L., 2000. Alpha-substituted N-(sulfonamido)alkyl-beta-aminotetralins: potent and selective neuropeptide Y Y5 receptor antagonists. J. Med. Chem. 43, 346–350.
- Zakrzewska, K.E., Sainsbury, A., Cusin, I., Rouru, J., Jeanrenaud, B., Rohner-Jeanrenaud, F., 1999. Selective dependence of intracerebroventricular neuropeptide Y-elicited effects on central glucocorticoids. Endocrinology 140, 3183–3187.
- Zammaretti, F., Panzica, G., Eva, C., 2001. Fasting, leptin treatment, and glucose administration differentially regulate Y(1) receptor gene expression in the hypothalamus of transgenic mice. Endocrinology 142, 3774–3782.
- Zarjevski, N., Cusin, I., Vettor, R., Rohner-Jeanrenaud, F., Jeanrenaud, B., 1993. Chronic intracerebroventricular neuropeptide-Y administration to normal rats mimics hormonal and metabolic changes of obesity. Endocrinology 133, 1753-1758.
- Zimanyi, I.A., Poindexter, G.S., 2000. NPY-ergic agents for the treatment of obesity. Drug Dev. Res. 51, 94-111.